

DIVISION OF MOLECULAR & GENOMIC PATHOLOGY

<i>(To Be Completed by MGP Staff)</i>					
Received Date		PHS #			
Solid Tumor Test Requisition					
PATIENT IDENTIFICATION					
Last Name		First Name		M.I.	SSN/MRN
Birthdate	Sex <input type="checkbox"/> M <input type="checkbox"/> F	Diagnosis	ICD-10 Code(s)		Surgical Path/Cytology #
CLIENT INFORMATION					
Requesting Institution/ Physician					
Requesting Physician Address					
Phone Number			Fax Number		
BILLING INFORMATION					
Person/Institution Responsible For Payment					
Billing Address					
Phone Number			Fax Number		
SPECIMEN INFORMATION * <i>SURGICAL PATHOLOGY/CYTOLOGY REPORT MUST BE INCLUDED</i>					
Collection Date:		Collection Time:		Source of Specimen:	
<input type="checkbox"/> Paraffin Sections		<input type="checkbox"/> Frozen Tissue		<input type="checkbox"/> Peripheral Blood	<input type="checkbox"/> Cyst Fluid
<input type="checkbox"/> FNA Fresh in Preservative Solution		<input type="checkbox"/> FNA Fixed		<input type="checkbox"/> Buccal Swab/Brush	<input type="checkbox"/> Bile Duct Brushing
TESTS					
BRAIN			THYROID		
<input type="checkbox"/> GlioSeq[®], Brain Tumors Mutation Panel: Mutations and CNVs in 30 genes (IDH1/2, BRAF, H3F3A, ATRX, SMO, etc.) and gene fusions (EGFRvIII, BRAF, FGFR3, etc.), NGS* <input type="checkbox"/> 1p/19q Deletion, LOH <input type="checkbox"/> 9p (CDKN2A,p16), LOH <input type="checkbox"/> 10q (PTEN), LOH <input type="checkbox"/> 17p (TP53), LOH <input type="checkbox"/> MGMT Methylation <input type="checkbox"/> IDH1 & IDH2 Mutation Analysis <input type="checkbox"/> BRAF Mutation Analysis <input type="checkbox"/> TERT Mutation Analysis (-124C>T, -146C>T) <input type="checkbox"/> EGFR Amplification, FISH			<input type="checkbox"/> ThyroSeq[®] v.3 GC, Thyroid Cancer Mutation Panel Mutations (112 genes for mutations, copy number alterations, gene fusions and gene expression), NGS* PANCREATIC CYSTS/ CHOLANGIO/ NEUROENDOCRINE <input type="checkbox"/> PancreaSeq[®], Pancreatic Cyst Fluid Panel (8 Genes), NGS* <input type="checkbox"/> BiliSeq, Bile Duct Brushing Panel (Mutations and CNVs in 28 genes, including KRAS, GNAS, TP53, PIK3CA, IDH1/2, etc.), NGS* <input type="checkbox"/> PanNeuroSeq, Neuroendocrine Tumors of the GI (Mutations and CNV in 10 genes including TP53, H3F3A, TERT, KRAS, ATRX, etc.), NGS*		
LUNG			COLON		
<input type="checkbox"/> Lung Panel (EGFR, KRAS, BRAF, ALK, MET mutations by NGS); ALK, RET, ROS1 fusion by FISH, MET amplification by FISH; PD- L1 expression by IHC. <input type="checkbox"/> OncoSeq Panel (mutations and CNVs in 28 genes, including EGFR, KRAS, BRAF, PIK3CA, MET, ERBB2, etc.) NGS <input type="checkbox"/> ALK, RET, ROS1 fusion, MET amplification by FISH <input type="checkbox"/> PD-L1 expression by IHC <input type="checkbox"/> EGFR Mutation Analysis (exons 18-21) <input type="checkbox"/> EGFR T790M Mutation Analysis (erlotinib resistance)			<input type="checkbox"/> OncoSeq Panel (28 genes for mutations and CNVs in colon cancer related genes, i.e. KRAS, NRAS, BRAF, PIK3CA, SMAD4, etc.), NGS* <input type="checkbox"/> Colon Panel (5 genes for mutations in colon cancer: KRAS, NRAS, HRAS, BRAF, and PIK3CA), NGS <input type="checkbox"/> KRAS Mutation Analysis (exons 1, 2) <input type="checkbox"/> BRAF Mutation Analysis (exon 15) <input type="checkbox"/> Microsatellite Instability (MSI)		
MELANOMA			OTHER		
<input type="checkbox"/> Melanoma panel (BRAF, NRAS, NF1), NGS <input type="checkbox"/> TERT Mutation Analysis (-124C>T, -146C>T)			<input type="checkbox"/> Personalized Cancer Mutation Panel (50 genes for mutations and CNVs), NGS* <input type="checkbox"/> Double primary testing for tumor de novo vs. metastasis <input type="checkbox"/> Tissue Identity <input type="checkbox"/> Other tests _____		

Specimen Instructions and Shipping Instructions

Formalin fixed and paraffin embedded tissue sections

- ◆ Tissue should be fixed in formalin and not exposed to decalcification solution. The paraffin block should contain no less than 3 mm area of tumor.
- ◆ Slides should be prepared by histology using a specific protocol for cutting molecular sections to avoid contamination of the tissue sections (available upon request).
- ◆ 1 H&E and 6 unstained sections are required for most of the tests. Ten unstained sections or more are required for some tests or if the tissue is small. Please call the lab if you have questions.
- ◆ Inclusion of normal patient tissue (either adjacent to tumor in the same block or separate block) is optimal for LOH and MSI analyses.
- ◆ Slides should be properly labeled with a block label that matches the surgical pathology specimen number on the surgical pathology report.
- ◆ Slides should be sent ambient temperature in proper storage containers (plastic slide boxes) to protect them during shipment.
- ◆ A surgical pathology and/or cytology report and completed requisition form must accompany all specimens.

Frozen or fresh tissue

- ◆ A minimum of 2 x 2 x 2 mm of frozen tissue is required; however, 5 x 5 x 5 mm is optimal.
- ◆ Collection date and time should be stated.
- ◆ Tissue specimen containing at least 50% of tumor cells can be either placed into cryogenic tube and snap frozen in liquid nitrogen, or placed into a tube with preservative solution provided by the Molecular & Genomic Pathology laboratory (request solution from the lab) and frozen at -20°C.
- ◆ Ship overnight on dry ice. A surgical pathology and/or cytology report and completed requisition form must accompany all specimens.

Fresh Fine Needle Aspiration (FNA) samples

- ◆ Fresh specimens should be collected into preservative solution provided by the Molecular & Genomic Pathology laboratory (request solution from the lab). Collection instruction will be provided with the solution.
- ◆ Collection date and time should be stated.
- ◆ Specimen can be refrigerated at 4°C for 12 hours or stored at -20°C prior to shipment.
- ◆ Ship with ice packs by overnight delivery. Shipping with dry ice is recommended for multiple tubes and when a delay in shipment is possible. A surgical pathology and/or cytology report and completed requisition form must accompany all specimens.

Formalin fixed and paraffin embedded Fine Needle Aspiration (FNA) samples

- ◆ 1 H&E and 10 unstained sections from cell block are required. A minimum of 300 tumor cells should be present on a slide. Please call the lab if you have any questions.
- ◆ Slides should be properly labeled with a number that matches the specimen number on the cytology report.
- ◆ Slides should be sent in proper storage containers (plastic slide boxes) to protect them during shipment. A surgical pathology and/or cytology report and completed requisition form must accompany all specimens.

Peripheral blood and bone marrow

- ◆ 2-5 ml of fresh peripheral blood collected in EDTA (purple top) tube or ACD (yellow top) tube.
- ◆ Blood should be refrigerated until shipment at 4°C.
- ◆ Shipment is at ambient temperature by overnight delivery in a properly labeled shipping container for biohazard substances. A surgical pathology and/or cytology report and completed requisition form must accompany all specimens.