

Myeloproliferative Neoplasm, CALR with Reflex to MPL, Varies

Overview

Useful For

Aiding in the distinction between a reactive cytosis and a myeloproliferative neoplasm when JAK2V617F testing result is negative

Reflex Tests

Test Id	Reporting Name	Available Separately	Always Performed
MPNML	MPL Exon 10 Sequencing,	No, (bill only)	No
	Reflex		

Testing Algorithm

This test reflexively evaluates for variants in the *CALR* and *MPL* genes commonly associated with *BCR/ABL1*-negative myeloproliferative neoplasms. The testing sequence is based on the reported frequency of gene variants in this disease group. It is usually ordered when a *JAK2* V617F result is known to be negative. Initial testing evaluates for the presence of the *CALR* insertions and deletions. If out-of-frame *CALR* insertions or deletions are detected, the testing algorithm ends. If the *CALR* result is negative or an in-frame *CALR* insertion or deletion is identified, then testing proceeds, at an additional charge, to evaluate for variants in exon 10 of the *MPL* gene by Sanger sequencing. An integrated report is issued with the summary of test results.

The following algorithms are available in Special Instructions: -Myeloproliferative Neoplasm: A Diagnostic Approach to Bone Marrow Evaluation

-Myeloproliferative Neoplasm: A Diagnostic Approach to Peripheral Blood Evaluation

Special Instructions

- <u>Myeloproliferative Neoplasm: A Diagnostic Approach to Peripheral Blood Evaluation</u>
- Myeloproliferative Neoplasm: A Diagnostic Approach to Bone Marrow Evaluation

Method Name

Polymerase Chain Reaction (PCR) and Fragment Analysis

NY State Available

Yes

Specimen

Specimen Type Varies



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Shipping Instructions

Specimen must arrive within 7 days of collection.

Necessary Information

The following information is required:

- 1. Pertinent clinical history
- 2. Clinical or morphologic suspicion
- 3. Date of collection
- 4. Specimen source

Specimen Required Submit only 1 of the following specimens:

Specimen Type: Blood

Container/Tube: Lavender top (EDTA) or yellow top (ACD-B)

Specimen Volume: 3 mL

Collection Instructions:

- 1. Invert several times to mix blood.
- 2. Send specimen in original tube.
- 3. Label specimen as blood.
- Specimen Stability Information: Ambient (preferred)/Refrigerate

Specimen Type: Bone marrow aspirate

Container/Tube: Lavender top (EDTA) or yellow top (ACD-B)

Specimen Volume: 2 mL

Collection Instructions:

- 1. Invert several times to mix specimen.
- 2. Send specimen in original tube.
- 3. Label specimen as bone marrow.
- Specimen Stability Information: Ambient (preferred)/Refrigerate
- **Specimen Type**: Extracted DNA from blood or bone marrow
- Container/Tube: 1.5 to 2 mL tube
- Specimen Volume: Entire specimen

Collection Instructions:

- 1. Indicate volume and concentration of DNA
- 2. Label specimen as extracted DNA from blood or bone marrow.
- Specimen Stability Information: Frozen (preferred)/Refrigerate/Ambient

Forms

If not ordering electronically, complete, print, and send a <u>Hematopathology/Cytogenetics Test Request</u> (T726) with the specimen.

Specimen Minimum Volume

Blood or Bone marrow: 0.5 mL



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Reject Due To

Gross	Reject
hemolysis	
Paraffin-embe	Reject
dded bone	
marrow	
aspirate clot or	
biopsy blocks	
Slides Paraffin	
shavings	
Moderately to	
severely	
clotted	

Specimen Stability Information

Specimen Type	Temperature	Time	Special Container
Varies	Varies	7 days	

Clinical & Interpretive

Clinical Information

JAK2 V617F variant is present in 95% to 98% of polycythemia vera (PV) patients, 50% to 60% of primary myelofibrosis (PMF) patients, and 50% to 60% of essential thrombocythemia (ET) patients. Detection of the *JAK2* V617F variant helps establish the diagnosis of a myeloproliferative neoplasm (MPN). However, a negative *JAK2* V617F result does not indicate the absence of MPN. Other important molecular markers in *BCR-ABL1*-negative MPN include *CALR* exon 9 variants (20%-30% of PMF and ET) and *MPL* exon 10 variants (5%-10% of PMF and 3%-5% of ET). Variants in *JAK2, CALR*, and *MPL* are essentially mutually exclusive. A *CALR* variant is associated with decreased risk of thrombosis in both ET and PMF, and confers a favorable clinical outcome in PMF patients. A triple negative (*JAK2* V617F, *CALR*, and *MPL*-negative) genotype is considered a high-risk molecular signature in PMF.

Reference Values

An interpretive report will be provided.

Interpretation

The results will be reported as 1 of the 3 following states: -Positive for CALR variant -Positive for MPL variant -Negative for CALR and MPL variants

Positive variants status is highly suggestive of a myeloid neoplasm and clinicopathologic correlation is necessary in all



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cases.

Negative variant status does not exclude the presence of a myeloproliferative neoplasm or other neoplasms.

Cautions

A positive result is not specific for a particular subtype of myeloproliferative neoplasm and clinicopathologic correlation is necessary in all cases.

A negative result does not exclude the presence of a myeloproliferative neoplasm or other neoplastic process.

Clinical Reference

1. Klampfl T, Gisslinger H, Harutyunyan AS, et al: Somatic mutation of calreticulin in myeloproliferative neoplasms. N Engl J Med 2013;369:2379-2390

2. Nangalia J, Massie CE, Baxter EJ, et al: Somatic CALR mutation in myeloproliferative neoplasms with nonmutated JAK2. N Engl J Med 2013;369:2391-2405

3. Rotunno G, Mannarelli C, Guglielmelli P, et al: Impact of calreticulin mutations on clinical and hematological phenotype and outcome in essential thrombocythemia. Blood 2014;123:1552-1555

4. Tefferi A, Lasho TL, Finke CM, et al: CALR vs JAK2 vs MPL-mutated or triple-negative myelofibrosis: clinical, cytogenetic and molecular comparisons. Leukemia advance online publication 21 January 2014

5. Pikman Y, Lee BH, Mercher T, et al: MPLW515L is a novel somatic activating mutation in myelofibrosis with myeloid metaplasia. FLoS Med 2006;3:e270

6. Pardanani A, Levine R, Lasho T, et al: *MPL*515 mutations in myeloproliferative and other myeloid disorders: a study of 1182 patients. Blood 2006;15:3472

Performance

Method Description

PCR amplification of *CALR* exon 9 is performed on DNA isolated from the patient sample. The PCR product is then run on an ABI Genetic Analyzer for fragment analysis to detect insertions and deletions. An unmutated *CALR* will show an amplicon at 266 bp, a mutated *CALR* with insertion will show an amplicon greater than 266 bp, and a mutated *CALR* with deletion will show an amplicon smaller than 266 bp. This assay has an analytical sensitivity of approximately 6% (ie, 6 variant-containing cells in 100 total cells) in most variant types, except for the rare type of 1-bp deletion, which has a sensitivity of approximately 20%. This is a laboratory developed test using analyte-specific reagents and research use only (RUO) reagents.(Unpublished Mayo method)

Genomic DNA is extracted and Sanger sequencing used to evaluate for variants in *MPL*, exon 10. The sensitivity of this assay is approximately 20%, such that samples containing lower percentages of mutated DNA will appear negative.(Unpublished Mayo method)

PDF Report

No

Day(s) Performed



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Monday through Friday

Report Available 7 to 10 days

Specimen Retention Time DNA: 3 months

Performing Laboratory Location Rochester

Fees & Codes

Fees

- Authorized users can sign in to <u>Test Prices</u> for detailed fee information.
- Clients without access to Test Prices can contact <u>Customer Service</u> 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact Customer Service.

Test Classification

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. It has not been cleared or approved by the US Food and Drug Administration.

CPT Code Information

81219-CALR (calreticulin) (eg, myeloproliferative disorders), gene analysis, common variants in exon 9 81339 -MPL (MPL proto-oncogene, thrombopoietin receptor) (eg, myeloproliferative disorder) gene analysis; sequence analysis, exon 10 (if appropriate)

LOINC[®] Information

Test ID	Test Order Name	Order LOINC [®] Value
MPNCM	MPN (CALR, MPL) Reflex	In Process

Result ID	Test Result Name	Result LOINC [®] Value
42393	MPNCM Reflex Result	82939-0
MP036	Specimen Type	31208-2
42392	Final Diagnosis	50398-7