



Test Definition: FB12

Vitamin B12 and Folate, Serum

Overview

Useful For

Investigation of macrocytic anemia

Workup of deficiencies seen in megaloblastic anemias

Investigation of suspected folate deficiency

Diagnosis of vitamin B12 deficiency-associated neuropathy

Profile Information

Test Id	Reporting Name	Available Separately	Always Performed
B12	Vitamin B12 Assay, S	Yes	Yes
FOL	Folate, S	Yes	Yes

Testing Algorithm

For more information, see [Vitamin B12 Deficiency Evaluation](#).

Special Instructions

- [Vitamin B12 Deficiency Evaluation](#)

Method Name

B12: Immunoenzymatic Assay

FOL: Competitive Binding Receptor Assay

NY State Available

No

Specimen

Specimen Type

Serum

Specimen Required

Patient Preparation:

1. **Fasting: 8 hours, required**
2. **Do not order** on patients who have recently received methotrexate or other folic acid antagonists.

Collection Container/Tube:

Preferred: Red top

Acceptable: Serum gel

Submission Container/Tube: Plastic vial

Specimen Volume: 1 mL serum

Collection Instructions: Centrifuge and aliquot serum into a plastic vial.

Forms

If not ordering electronically, complete, print, and send 1 of the following forms with the specimen:

[-Kidney Transplant Test Request](#)

[-Benign Hematology Test Request Form \(T755\)](#)

Specimen Minimum Volume

Serum: 0.5 mL

Reject Due To

Gross hemolysis	Reject
Gross lipemia	OK

Specimen Stability Information

Specimen Type	Temperature	Time	Special Container
Serum	Refrigerated (preferred)	7 days	
	Frozen	90 days	

Clinical & Interpretive

Clinical Information

Vitamin B12:

Vitamin B12 (cobalamin) is necessary for hematopoiesis and normal neuronal function. In humans, it is obtained only from animal proteins and requires intrinsic factor (IF) for absorption. The body uses its vitamin B12 stores very economically, reabsorbing vitamin B12 from the ileum and returning it to the liver; very little is excreted.

Vitamin B12 deficiency may be due to lack of IF secretion by gastric mucosa (eg, gastrectomy, gastric atrophy) or intestinal malabsorption (eg, ileal resection, small intestinal diseases).

Vitamin B12 deficiency frequently causes macrocytic anemia, glossitis, peripheral neuropathy, weakness, hyperreflexia, ataxia, loss of proprioception, poor coordination, and affective behavioral changes. These manifestations may occur in any combination; many patients have neurologic defects without macrocytic anemia.

Pernicious anemia is a macrocytic anemia caused by vitamin B12 deficiency that is due to a lack of IF secretion by gastric mucosa.

Serum methylmalonic acid and homocysteine levels are also elevated in vitamin B12 deficiency states.

Folate:

The term folate refers to all derivatives of folic acid. For practical purposes, serum folate is almost entirely in the form of *N*-(5)-methyl tetrahydrofolate.(1)

Approximately 20% of the folate absorbed daily is derived from dietary sources; the remainder is synthesized by intestinal microorganisms. Serum folate levels typically fall within a few days after dietary folate intake is reduced and may be low in the presence of normal tissue stores. Red blood cell folate levels are less subject to short-term dietary changes.

Significant folate deficiency is characteristically associated with macrocytosis and megaloblastic anemia. Lower than normal serum folate also has been reported in patients with neuropsychiatric disorders, in pregnant women whose fetuses have neural tube defects, and in women who have recently had spontaneous abortions.(2) Folate deficiency is most commonly due to insufficient dietary intake and is most frequently encountered in pregnant women or in alcoholics.

Other causes of low serum folate concentration include:

- Excessive utilization (eg, liver disease, hemolytic disorders, and malignancies)
- Rare inborn errors of metabolism (eg, dihydrofolate reductase deficiency, forminotransferase deficiency, 5,10-methylenetetra-hydrofolate reductase deficiency, and tetrahydrofolate methyltransferase deficiency)

Reference Values**VITAMIN B12**

180-914 ng/L

FOLATE

> or =4.0 mcg/L

<4.0 mcg/L suggests folate deficiency

Interpretation**Vitamin B12:**

Concentration of vitamin B12 less than 180 ng/L may cause megaloblastic anemia and/or peripheral neuropathies.

Vitamin B12 concentrations less than 150 ng/L are considered evidence of vitamin B12 deficiency.

Vitamin B12 concentrations between 150 ng/L and 400 ng/L are considered borderline.

Follow-up testing for antibodies to intrinsic factor (IF) (IFBA / Intrinsic Factor Blocking Antibody, Serum) is recommended to identify this potential cause of vitamin B12 malabsorption.

For specimens without antibodies, follow-up testing of vitamin B12 tissue deficiency by measuring methylmalonic acid (MMA) (MMAS / Methylmalonic Acid, Quantitative, Serum) and/or homocysteine (HCYSP / Homocysteine, Total, Plasma) may be indicated if the patient is symptomatic.

A normal serum concentration of vitamin B12 does not rule out tissue deficiency of vitamin B12. The most sensitive test for vitamin B12 deficiency at the cellular level is the assay for MMA. If clinical symptoms suggest deficiency,

measurement of MMA and homocysteine should be considered, even if serum vitamin B12 concentrations are normal.

Folate:

Serum folate is a relatively nonspecific test.(1) Low serum folate levels may be seen in the absence of deficiency and normal levels may be seen in patients with macrocytic anemia, dementia, neuropsychiatric disorders, and pregnancy disorders.

Results less than 4 mcg/L are suggestive of folate deficiency. The cut-off is based on consensus and was derived from the US NHANES III data.(2)

Evaluation of macrocytic anemias commonly requires measurement of the serum concentration of both vitamin B12 and folate; ideally they should be measured at the same point in time.

Additional testing with homocysteine and MMA determinations may help distinguish between B12 and folate deficiency states. In folate deficiency, homocysteine levels are elevated and MMA levels are normal. In vitamin B12 deficiency, both homocysteine levels and MMA levels are elevated.

For more information, see [Vitamin B12 Deficiency Evaluation](#).

Cautions**Vitamin B12:**

Patients taking vitamin B12 supplementation may have misleading results.

Many other conditions are known to cause an increase or decrease in serum vitamin B12 concentration.

Conditions known to cause increases:

- Ingestion of vitamin C
- Ingestion of estrogens
- Ingestion of vitamin A
- Hepatocellular injury
- Myeloproliferative disorder
- Uremia

Conditions known to cause decreases:

- Pregnancy
- Aspirin
- Anticonvulsants
- Ingestion of ethanol
- Contraceptive hormones
- Smoking
- Hemodialysis
- Multiple myeloma

The evaluation of macrocytic anemia requires measurement of both vitamin B12 and folate levels; ideally they should be measured simultaneously.

Some patients who have been exposed to animal antigens, either in the environment or as part of treatment or imaging

procedure, may have circulating anti-animal antibodies present. These antibodies may interfere with the assay reagents to produce unreliable results.

Folate:

Patients with combined deficiency of folate and iron may not demonstrate the erythrocyte macrocytosis that is typical of folate deficiency anemia. In these patients, however, the red cell distribution width will typically be elevated.

Nonfasting specimens yield falsely elevated results.

Recent folic acid administration or dietary folate intake could result in normal or elevated values and possibly mask an underlying folate deficiency.

Patients taking folate may have misleading results.

Folates other than *N*-(5)-methyltetrahydrofolate and folic acid antagonists (such as methotrexate) may, under some circumstances, be present in serum and will also be measured by this method.

Serum folate measurement is preferred over red blood cell (RBC) folate measurement due to considerable analytic variability (coefficient of variation; CV) of assays. Both results give the same interpretation (internal Mayo study), therefore RBC folate quantitation is not recommended. Additional serum testing with homocysteine and methylmalonic acid (MMA) determinations may help distinguish between vitamin B12 and folate deficiency states. In folate deficiency, homocysteine levels are elevated and MMA levels are normal. In vitamin B12 deficiency, the analytic variability CV of both serum and RBC folate assays is considerable. Homocysteine and MMA levels are alternate determinates of folate deficiency.

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Supportive Data

See Individual Unit Codes

Clinical Reference

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Performance

Method Description

Vitamin B12:

The Access Vitamin B12 assay is a competitive-binding immunoenzymatic assay performed on the Beckman Coulter DXI 800. A sample is added to a reaction vessel along with alkaline potassium cyanide and dithiothreitol. This treatment denatures vitamin B12 binding proteins and converts all forms of vitamin B12 to the cyanocobalamin form. After neutralization, intrinsic factor-alkaline phosphatase conjugate and paramagnetic particles coated with goat antimouse IgG:mouse monoclonal anti-intrinsic factor are added to the sample. Vitamin B12 in the sample binds to the intrinsic factor conjugate, preventing the conjugate from binding to the solid phase anti-intrinsic factor. After incubation in a reaction vessel, materials bound to the solid phase are held in a magnetic field, while unbound materials are washed away. The chemiluminescent substrate Lumi-Phos 530 is added to the vessel and light generated by the reaction is measured with a luminometer. The photon production is inversely proportional to the concentration of vitamin B12 in the sample. The amount of analyte in the sample is determined by means of a stored, multipoint calibration curve.(Package insert: Access Vitamin B12. Beckman Coulter, Inc.; C64657F, 03/2025)

Folate:

The Access Folate assay is a competitive-binding receptor assay performed on the Beckman Coulter DXI 800. A serum sample is treated to release folate from endogenous binding proteins. After neutralization of the reaction mixture, folate-binding protein, mouse antifolate-binding protein, folic acid-alkaline phosphatase conjugate, and goat antimouse capture antibody coupled to paramagnetic particles are added to the reaction vessel. Folate in the sample competes with the folic acid-alkaline phosphatase conjugate for binding sites on a limited amount of folate-binding protein. Resulting complexes bind to the solid phase via mouse antifolate binding protein. After incubation in a reaction vessel,

materials bound to the solid phase are held in a magnetic field, while unbound materials are washed away. The chemiluminescent substrate Lumi-Phos 530 is added to the vessel and light generated by the reaction is measured with a luminometer. The light production is inversely proportional to the concentration of folate in the sample. The amount of analyte in the sample is determined from a stored, multipoint calibration curve. The assay is standardized to the World Health Organization (WHO) International Standard 03/178. (Package insert: Access Folate. Beckman Coulter, Inc.; B03897J, 08/2024)

PDF Report

No

Day(s) Performed

Monday through Friday, Sunday

Report Available

1 to 3 days

Specimen Retention Time

14 days

Performing Laboratory Location

Mayo Clinic Jacksonville Clinical Lab

Fees & Codes**Fees**

- Authorized users can sign in to [Test Prices](#) for detailed fee information.
- Clients without access to Test Prices can contact [Customer Service](#) 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact [Customer Service](#).

Test Classification

This test has been cleared, approved, or is exempt by the US Food and Drug Administration and is used per manufacturer's instructions. Performance characteristics were verified by Mayo Clinic in a manner consistent with CLIA requirements.

CPT Code Information

82607-Vitamin B12

82746-Folate

LOINC® Information

Test ID	Test Order Name	Order LOINC® Value
FB12	Vitamin B12 and Folate, S	96805-7

Result ID	Test Result Name	Result LOINC® Value
B12	Vitamin B12 Assay, S	2132-9

FOL	Folate, S	2284-8
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