

# **Test Definition: LCJC**

JC Virus, Molecular Detection, PCR, Spinal Fluid

#### Overview

#### **Useful For**

Aiding in diagnosing progressive multifocal leukoencephalopathy due to JC virus

This test is **not to be used** as a diagnostic tool for Creutzfeldt-Jakob disease.

This test is not recommended for screening asymptomatic patients

#### **Testing Algorithm**

For more information see Meningitis/Encephalitis Panel Algorithm

#### **Special Instructions**

Meningitis/Encephalitis Panel Algorithm

#### Method Name

Real-Time Polymerase Chain Reaction (PCR)/DNA Probe Hybridization

NY State Available

#### Specimen

# Specimen Type

CSF

#### Specimen Required

Supplies: Sarstedt Aliquot Tube, 5mL (T914) Container/Tube: Preferred: Aliquot tube Acceptable: Sterile screw cap vial Specimen Volume: 0.5 mL Collection Instructions: Do not centrifuge.

#### **Specimen Minimum Volume**

0.3 mL

#### Reject Due To

Heat-inactivate	Reject
d specimens	



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#### **Specimen Stability Information**

Specimen Type	Temperature	Time	Special Container
CSF	Refrigerated (preferred)	7 days	
	Frozen	7 days	

### **Clinical & Interpretive**

#### **Clinical Information**

JC virus (JCV), a member of the genus *Polyomavirus*, is a small nonenveloped DNA-containing virus. Primary infection occurs in early childhood, with a prevalence of greater than 80%.(1) The virus is latent but can reactivate in immunosuppressed patients, especially those with AIDS.

JCV is recognized as the etiologic agent of progressive multifocal leukoencephalopathy (PML), a fatal demyelinating disease of the central nervous system.(2,3) Histologic examination of brain biopsy tissue may reveal characteristic pathologic changes localized mainly in oligodendrocytes and astrocytes. Detection of JCV DNA by polymerase chain reaction (PCR) (target gene, large T antigen) in the cerebrospinal fluid specimens of patients with suspected PML infection has replaced the need for biopsy tissue for laboratory diagnosis.(4) Importantly, the PCR test is specific with no cross-reaction with BK virus, a closely related polyomavirus.

#### Reference Values

Negative Reference values apply to all ages.

#### Interpretation

Detection of JC virus (JCV) DNA supports the clinical diagnosis of progressive multifocal leukoencephalopathy due to JCV.

#### Cautions

A negative result does not rule out the possibility of JC virus (JCV) infection.

The reference value in cerebrospinal fluid is "negative" for this assay, although JCV DNA may be detectable in the absence of clinical symptoms in certain patient populations.(5,6) However, this assay is only to be used for patients with appropriate neurological and neuroradiological features of progressive multifocal leukoencephalopathy, and is not indicated for screening asymptomatic patients.

#### Supportive Data

The following data supports the use of this assay for clinical testing.

#### Accuracy:

Twenty-six negative cerebrospinal fluid (CSF) specimens were spiked with JC virus (JCV)-positive control plasmid at the limit of detection (approximately 10 targets/mcL). The 26 spiked specimens were run in a blinded manner with 14 negative (nonspiked) specimens. 100% of the spiked specimens were positive and 100% of the nonspiked specimens were negative.

Analytical Sensitivity/Limit of Detection (LoD):



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The lower limit of detection (LoD) of this assay is 10 DNA target copies per mcL in CSF.

Analytical Specificity:

No PCR signal was obtained from the extracts of 15 viral isolates that may cause similar symptoms or be found in the CSF, including herpes simples virus (HSV) types 1 and 2, Epstein-Barr virus (EBV), cytomegalovirus (CMV), human herpesvirus (HHV)-6, HHV-7, HHV-8, enterovirus, mumps, adenovirus, BK virus and Simian virus 40 (SV40).

Precision:

Interassay precision was 100% and intraassay precision was 100%.

Reference Range:

The reference range in CSF is "negative" for this assay.

Reportable Range:

This is a qualitative assay and the results are reported as either negative or positive for targeted JCV DNA.

#### **Clinical Reference**

1. Safak M, Khalili K: An overview: human polyomavirus JC virus and its associated disorders. J Neurovirol. 2006;9 Suppl 1:3-9. doi: 10.1080/13550280390195360

2. Khalili K, White MK: Human demyelinating disease and the polyomavirus JCV. Mult Scler. 2006 Apr;12(2):133-142

3. Ahsan N, Shah KV: Polyomaviruses and human diseases. Adv Exp Med Bio.l 2006;577:1-18. doi:

10.1007/0-387-32957-9\_1

4. Romero JR, Kimberlin DW: Molecular diagnosis of viral infections of the central nervous system. Clin Lab Med. 2003 Dec;23(4):843-865

5. Chen Y, Bord E, Tompkins T, et al: Asymptomatic reactivation of JC virus in patients treated with natalizumab. N Engl J Med. 2009 Sep 10;361(11):1067-1074

6. Egli A, Infanti L, Dumoulin A, et al: Prevalence of polyomavirus BK and JC infection and replication in 400 healthy donors. J Infect Dis. 2009 Mar 15;199(6):837-846

7. Tan CS, Koralnik IJ: JC, BK, and other Polyomaviruses: Progressive multifocal leukoencephalopathy (PML). In: Bennett JE, Dolin R, Blaser MJ, eds. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases. 9th ed. Elsevier; 2020:1931-1939

# Performance

#### **Method Description**

Viral nucleic acid is extracted from the specimen using the MagNA Pure automated instrument (Roche Applied Science). Primers are directed to the large T antigen gene, which is a conserved sequence specific for JC virus (JCV). This assay detects only JCV; it does not detect BK Virus or Simian Virus 40 (SV40) (other polyomaviruses). The LightCycler instrument (Roche Applied Science) amplifies and monitors the development of target nucleic acid sequences after the annealing step during PCR cycling. This automated PCR system can rapidly detect amplicon development through stringent air-controlled temperature cycling in capillary cuvettes. The detection of amplified products is based on the fluorescence resonance energy transfer (FRET) principle. For FRET product detection, a hybridization probe with a donor fluorophore, fluorescein, on the 3'-end is excited by an external light source and emits light that is absorbed by a second



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hybridization probe with an acceptor fluorophore, LC-Red 640, at the 5'-end. The acceptor fluorophore then emits a light of a different wavelength that can be measured with a signal that is proportional to the amount of specific PCR product.(Unpublished Mayo method)

#### PDF Report

No

Day(s) Performed Monday through Saturday

Report Available

2 to 5 days

**Specimen Retention Time** 14 days

### **Performing Laboratory Location**

Jacksonville

# Fees & Codes

#### Fees

- Authorized users can sign in to <u>Test Prices</u> for detailed fee information.
- Clients without access to Test Prices can contact <u>Customer Service</u> 24 hours a day, seven days a week.
- Prospective clients should contact their account representative. For assistance, contact Customer Service.

# **Test Classification**

This test was developed and its performance characteristics determined by Mayo Clinic in a manner consistent with CLIA requirements. It has not been cleared or approved by the US Food and Drug Administration.

#### **CPT Code Information**

87798

#### LOINC<sup>®</sup> Information

Test ID	Test Order Name	Order LOINC <sup>®</sup> Value
LCJC	JC Virus PCR, CSF	33295-7

Result ID	Test Result Name	Result LOINC <sup>®</sup> Value
88909	JC Virus PCR, CSF	33295-7